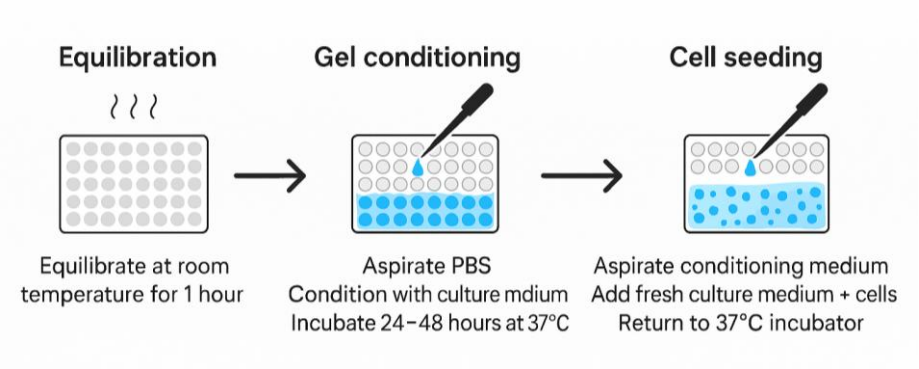
PROTOCOL

LifeGel Plates preparation for cell-based assays.

Materials required:
- 48-, 96- or 384-well LifeGel plate [RRLG96; RRLG48; RRLG384]
- Pipette/multichannel pipettor
- Complete culture medium

1. **Equilibration**
   Remove LifeGel plates from the refrigerator 24 or 48 hours before seeding. Allow them to equilibrate at room temperature for approximately 1 hour.
2. **Gel conditioning**
   Aspirate the PBS from each well*. Add 400 µl of culture medium per well for 48-well LifeGel plates, 150µl for 96-well LifeGel plates or 60 µl for 384-well plates to condition the gel. Incubate the plates at 37°C in a humidified incubator.
3. **Cell seeding**
   After 24 or 48 hours of conditioning, remove the plates from the incubator. Aspirate the conditioning medium from the wells*.
   Add 400µl (48-well), 150 µl (96-well) or 60 µl (384-well) of fresh culture medium containing cell suspension to each well.
   Return the seeded LifeGel plates to the 37°C incubator for cell culture.



* Be careful when aspirating PBS or conditioning medium to avoid disturbing the gel.

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